



Author: M.P. Field<sup>1</sup>, T.M. Conway<sup>2,3</sup>, B.A. Summers<sup>2</sup>, N. Saetveit<sup>1</sup>, and J.C. Sakowski<sup>1</sup>

## Automated Processing of Seawater Samples for Iron Isotope Ratio Determination

Poster Presented at Goldschmidt in Barcelona, Spain 2019

### Introduction

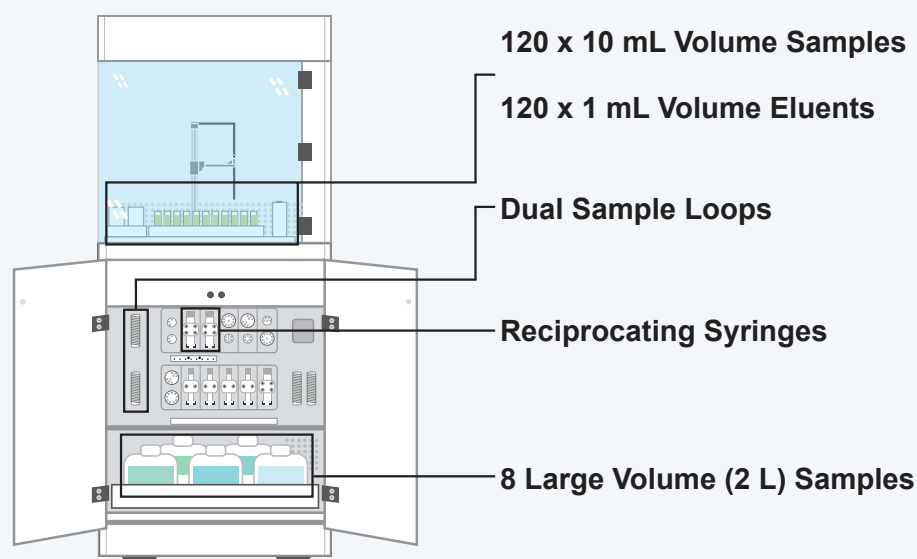
Bioactive metals, such as iron, act as important nutrients in the ocean and are often present at very low concentrations, which limit phytoplankton growth. In recent years, measurement of the dissolved stable isotope ratio of bioactive metals in seawater has provided insight into the biogeochemical cycling of these elements. Iron isotope ratios, in particular, have been shown to be useful tracers of iron sources across the open oceans [1].

High precision isotope ratio analysis requires sample amounts ranging from 2 to >100 ng, depending on the element and isotopes of interest. Trace and bioactive metals at low natural concentrations (1-100 ng L<sup>-1</sup>) in seawater pose a significant challenge with respect to sample volume requirements (~1-4 L), the seasalt matrix, and sample processing. Currently, two-step manual processes are employed to first preconcentrate a suite of metals in batch mode and then purify (chromatographically isolate) the metal(s) of interest for isotopic analysis (e.g. [2]), with methods often only optimized for 1 or 2 elements at a time.

Here we investigate a fully automated complete sample processing procedure using a newly developed, large-volume (up to 2 L), seawater preconcentration system in conjunction with a commercially available chromatography system (prepFAST MC). The first automation system uses a 200  $\mu$ L seaFAST column to perform a 1000x (400 mL to 400  $\mu$ L) preconcentration/matrix removal step for a suite of transition row and REEs. The prepFAST MC then purifies discrete Cu, Fe, and Zn fractions for isotopic analysis.

Elemental concentrations determined using HR-ICP-MS (Element 2) are used to calculate the total procedural blank, recovery, and carryover. Fe isotope ratios for seawater samples processed by both methods are measured using a HR-MC-ICP-MS (Neptune Plus) and compared for precision and accuracy with samples measured using a published batch extraction and column purification method [2].

## Step 1 - seaFAST Litre Preconcentrates Trace Metals from Seawater



## Experimental Conditions

- Seawater sample from 3000 m depth, Tropical Western South Atlantic Ocean
- Sample Volume 370 mL
- 200  $\mu$ L seaFAST column
- 600  $\mu$ L 10M HCl Eluent
- 120 min/sample

[illegible]

### Vial+1M HCl dry down Blank

- Cu: 11 pg
- Fe: 23 pg
- Zn: 19 pg

## seaFAST Litre Blank\*

- Cu: 38 pg
- Fe: 134 pg
- Zn: 34 pg

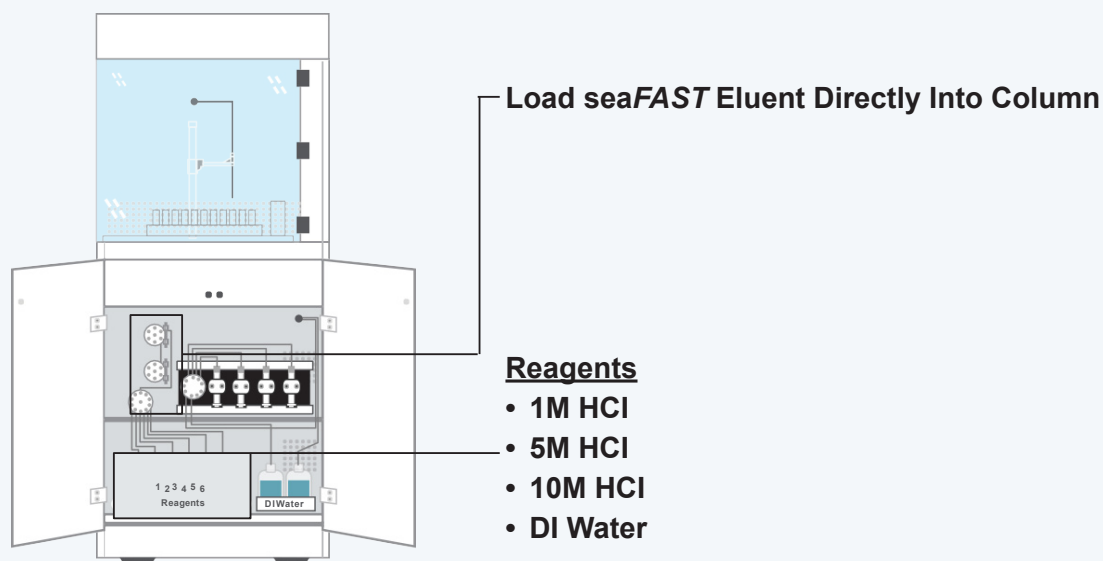
## seaFAST Litre Recovery\*

- Cu: 100.1%
- Fe: 99.8%
- Zn: 98.4%

\*Average of 3 samples

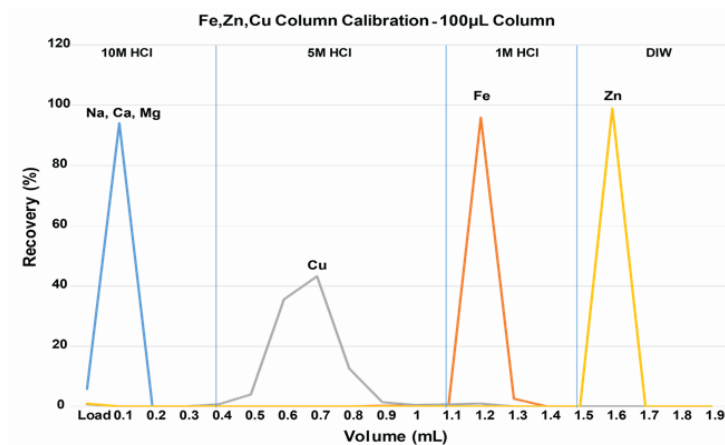


## Step 2 - prepFAST MC Purifies Fractions for Isotopic Analysis



### Experimental Conditions

- Preconcentrated seawater sample eluted in 10M HCl
- Load 600 mL
- 100  $\mu$ L Fe Zn Cu Column (p/n CF-MC-FeZnCu-0100)
- Load 10M HCl
- Wash 10M HCl
- Elute Cu, Fe and Zn fractions



### prepFAST MC Blank\*

- Cu: 7 pg
- Fe: 119 pg
- Zn: 19 pg

### prepFAST MC Recovery\*

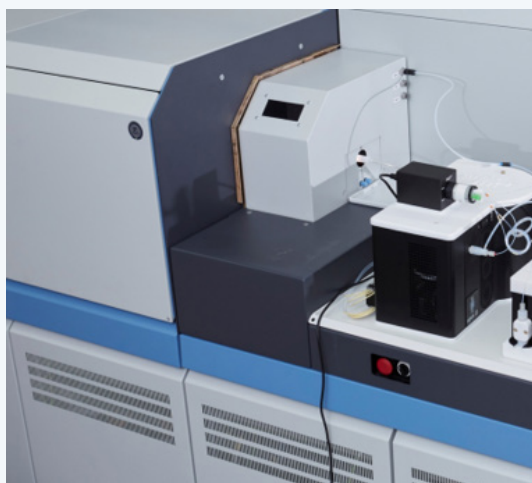
- Cu: 92.8%
- Fe: 99.5%
- Zn: 102.5%

### prepFAST MC Carryover\*

- Cu: 500x
- Fe: 400x
- Zn: 1500x

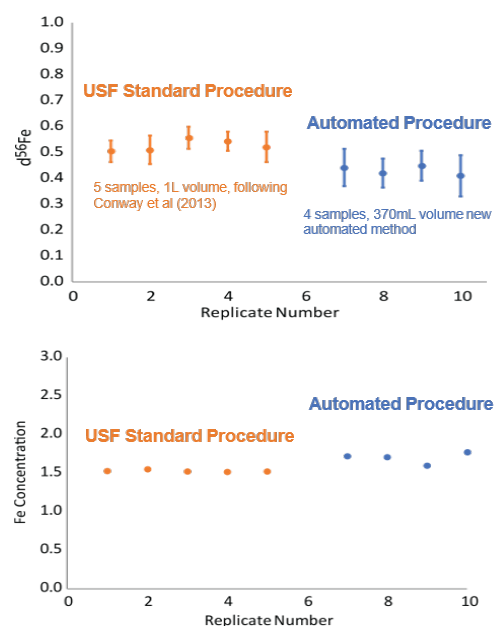
\*Average of 3 samples

### Step 3 - apex $\Omega$ Neptune Plus Analyze Sample for Precise Isotopic Ratios



#### Experimental Conditions

- Neptune Plus with Ni Jet sampler and AI X skimmer cone
- Apex  $\Omega$  with ~5 L Ar, no N.
- ~120  $\mu\text{L}/\text{min}$  ESI PFA neb
- Spiked 1:2 with a 57-58 Fe double spike from isoflex
- Data reduction through typical double-spike calculation following Siebert et al.
- Long-term analytical precision based on repeated analysis of the NIST 3126a standard of  $0.36 \pm 0.04$  permil based on 190 measurements over 15 runs over 2 years.



#### Conclusions

A novel automated procedure for purification of Cu, Fe and Zn from seawater has been developed and tested for Fe isotope accuracy and precision. The automated method has:

- High recovery
- Low blank
- Low carryover

#### References

1. Conway and John, (2014), *Nature*, 511, 212-215.
2. Conway et al., (2013), *ACA*, 793, 44-52.